

A Current Update on the Use of Alpha Lipoic Acid in the Management of Type 2 Diabetes Mellitus

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Abstract: Type 2 Diabetes Mellitus (T2DM) which is characterised by insulin resistance, is closely linked to the triad of glucolipotoxicity, inflammation and oxidative stress. Increased adiposity, leading to increased free fatty acids (FFAs), contributes to insulin resistance by disrupting the signal transduction pathway of insulin mediated glucose disposal, and causes impaired insulin secretion. Hyperglycaemia and dyslipidaemia driven oxidative stress resulting from enhanced free-radical formation and/or defects in antioxidant defence is implicated in the pathogenesis of diabetic neuropathy (DN). This and other inflammatory pathways account for a complex network of interacting metabolic factors responsible for causing diabetes and her complications. There is growing evidence that Alpha Lipoic Acid (ALA) has beneficial effects on the treatment of T2DM and some of its complications. It represents an attractive pharmacological target in the treatment of T2DM by modulating the signal transduction pathways in insulin resistance and antagonizing the oxidative and inflammatory stresses, which are major players in the pathogenesis of this disorder. A potent anti-oxidant and free radical scavenger, ALA also targets cellular signal transduction pathways which increases glucose uptake and utilization, thus providing specific targeted therapy in the treatment of insulin resistance and diabetic neuropathy. Apart from the rare risk of Insulin Autoimmune Syndrome (IAS), ALA has shown to be relatively safe, even in patients with renal and liver failure. This review focuses and summarises the molecular mechanisms of T2DM, and underlines the therapeutic value of ALA in this globally significant disease.

Key Words: ALA, anti-oxidant, diabetes, neuropathy, radical, insulin resistance.

INTRODUCTION

Alpha Lipoic Acid (ALA), also known as thiotic acid in Europe, is an 8-carbon, cyclic disulfide thio-octanoic acid that was first described by Lester in 1951 [1].

Having an asymmetric carbon atom, ALA has two isomers, R-enantiomer (R-LA) and S-enantiomer (S-LA). Only the R-isomer is endogenously synthesized and bound to protein. R-LA, being an essential cofactor for the biogenesis of mitochondrial enzymes, is found as a naturally occurring prosthetic group in the alpha-keto acid and pyruvate dehydrogenase complexes and is both easily taken up into cells, as well as being a fundamental player in metabolism [2-4].

ROLE IN OXIDATIVE STRESS

Oxidative stress, characterized by a persistent imbalance between the production of highly reactive- reactive oxygen species (ROS) and reactive nitrogen species (RNS) and anti-oxidant defences, leads to an altered cellular redox status and subsequent tissue damage [5, 6]. It was defined by Seis as a change in the pro-oxidant/anti-oxidant balance, in favour of the former, potentially leading to biological damage [7]. This biological damage, which includes DNA mutations and modifications to proteins and lipids, has been shown to be responsible for many modern diseases like cancer, cardiovascular disease, diabetes mellitus and even ageing.

The free radical theory of ageing by Harman introduced the idea that endogenous free radicals produced by cells

results in the accumulation of cellular damage [8, 9]. However, it was Ozawa's theory of redox mechanism of mitochondrial ageing which highlighted the importance of the mitochondria as an important source of ROS which usually occurs as a result of electron leakage in complexes I and III of the electron transport chain [10]. Mitochondria DNA has no histone protection or efficient repair system and is prone to the accumulation of DNA damage, which further lowers the bioenergetic capacity of mitochondria, resulting in a vicious cycle of increasing ROS production [11, 12].

While anti-oxidants serve to limit this damage, ALA contributes to the defence against oxidative stress by increasing the synthesis of anti-oxidants like glutathione, one of the most abundant intra-cellular anti-oxidants in the body [13, 14]. There is also evidence that ALA has the ability to directly scavenge ROS and RNS. Furthermore, the reduced form of ALA, dihydrolipoic acid (DHLA), *via* its reducing property, has the ability to reduce the oxidized form of other anti-oxidants such as ascorbic acid, alpha-tocopherol and coenzyme Q10 [15-17]. Hence, ALA has the theoretical potential to 'perpetuate' its antioxidant effect even after it is cleared systemically from the plasma, and thus plays a vital role in the regulation of other antioxidants and the synergism of antioxidants as a whole, described by Packer *et al.* [15] as the body's "antioxidant network". In addition, other yet unknown downstream actions of ALA with effects on this network may persist long after ALA has been cleared from the system.

Finally, ALA possesses metal chelating capacities which has been shown to reduce iron and copper mediated oxidative damage *in vitro* [18, 19], and reverse age-related iron

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accumulation and anti-oxidant depletion in the rat cerebral cortex [20]. Hence, the ability of ALA to alter the redox status of cells and interact with other antioxidants provides an attractive therapeutic potential in conditions where oxidative stress is involved [21].

ROLE IN SIGNAL TRANSDUCTION

Another important role of ALA is in the area of signal transduction. ALA acutely activates insulin receptors (IR) in its reduced state, which leads to a cascade of substrate phosphorylation that causes the translocation of glucose transporters (GLUT) from the cytoplasm to the cell surface. This results in peripheral glucose disposal, and a defect at any point in this pathway can lead to insulin resistance [22, 23].

ALA is also involved in the regulation of the nuclear factor-kappa B (NF-kB) and Akt signaling pathways [24]. NF-kB has been shown to regulate genes related to inflammation and cell cycle control, which have been implicated in the development of atherosclerosis, insulin resistance, pancreatic β cell cycle and even increasing chemosensitivity of mitotic lesions [25]. Results of cell line studies have shown that physiologic amounts of ALA inhibit NF-kB nuclear translocation, thus preventing its downstream effects on target gene expression [26]. ALA offers the potential for improved molecular therapeutic strategies.

PHARMACOKINETICS

ALA is soluble in both water and organic solvents [15]. A few studies demonstrated that up to 30-40% of orally ingested racemic ALA is absorbed with peak plasma concentrations higher in the R-isomer compared to the S-isomer, suggesting better absorption in the former as well as in a fasted state [27, 28].

That ALA is both soluble in water and organic solvents is significant, because although studies have shown no significant tissue accumulation and free plasma ALA levels after an oral ingestion is almost negligible due to rapid clearance rate, ALA is able to exert its powerful antioxidant effects because of this unique amphiphilic property. Because it is both hydrophobic and hydrophilic, it is able to elicit antioxidant effects by scavenging ROS and RNS in both media of cells and blood i.e. hydrophilic (cytoplasm and serum) and hydrophobic (plasma membrane and lipoprotein) [29]. Boreca *et al.* demonstrated a decrease in oxidative stress in diabetic patients on ALA treatment compared to those that were not, by measuring the plasma lipid hyperoxide: (alpha tocopherol/cholesterol) ratio. This was found to be independent of poor glycaemic control and albuminuria [30].

Relatively little is known regarding the excretion of ALA, except that renal excretion does not appear to play a significant role, and it has been shown to be safe in patients with End-Stage Renal Failure (ESRF) as well as in liver disease [31-33]. In fact, it has been used in the treatment of chronic hepatitis C, metal intoxication and liver damage secondary to alcohol, Amanita mushrooms [34], and carbon tetrachloride (CCl₄) with encouraging results [35].

PATHOGENESIS OF TYPE 2 DIABETES MELLITUS

Type 2 Diabetes Mellitus (T2DM) especially in the early stages, is characterized by insulin resistance secondary to the

triad of glucolipotoxicity, inflammation and oxidative stress. Increased adiposity, thought to be one of the initiating events, results in increased free fatty acid (FFA) release. Increased FFAs then contributes to insulin resistance by disrupting the signal transduction pathway of insulin mediated glucose disposal as well as cause impaired insulin secretion through pancreatic beta cell apoptosis. FFAs decrease insulin sensitivity through the induction of oxidative stress and the activation the serine kinases, protein kinase C α (PKC α) and inhibitor kappa b kinase (IKK β), which are known mediators of inflammation [36]. In fact, both chronic over-nutrition (obesity) and T2DM may represent pro-inflammatory states as increased concentrations of tumour necrosis factor-alpha (TNF- α) in these conditions have been found to interfere with insulin action by suppressing insulin signal transduction [37, 38].

Compelling evidence from both the United Kingdom Prospective Diabetes Study (UKPDS) and Diabetes Control and Complications Trial (DCCT) has shown that chronic hyperglycemia significantly increases the risks of developing complications such as nephropathy, neuropathy and retinopathy and tight glycaemic control is critical for the prevention of these complications [39]. One of the ways chronic hyperglycaemia contributes to the development of diabetic complications is through increased cellular oxidative stress. Diabetic neuropathy (DN) is one such example where the increased conversion of glucose to sorbitol *via* the sorbitol-aldose reductase pathway (polyol pathway) by aldose reductase results in an accumulation of intracellular sorbitol [40]. It is also associated with decreased myo-inositol due to a myo-inositol-related defect in nerve Na(+)-K(+)-ATPase, and decreased taurine levels (mechanism unclear) [41, 42]. The outcome of this metabolic alteration is a decreased NA+K+ ATPase activity, which in turns slows nerve conductivity [43]. Hence, in addition to hyperglycaemia and dyslipidaemia driven oxidative stress, the presence of polyol pathway activation and advanced glycated end products (AGEs), coupled to protein kinase C (PKC) activation account for the already complex network of interacting metabolic factors responsible for the endothelial dysfunction and reduced nerve perfusion and function [44].

Another way in which hyperglycaemia increases the risk of cellular oxidative damage is through a reduction in antioxidant defence [45], which has been observed during oral glucose challenge [46, 47]. Moreover, the conversion of glucose to sorbitol, and later fructose, results in the depletion of cellular nicotinamide adenine dinucleotide phosphate (NADPH) and oxidized nicotinamide adenine dinucleotide (NAD⁺) stores. This increases the vulnerability of the cell to the damaging actions of ROS. Furthermore, hyperglycaemia promotes the formation of ROS [48-50] by the auto-oxidation of glucose *via* the mitochondrial electron transport chain [51] and the formation of AGEs [52, 53].

In summary, hyperglycemia and increased FFA favours the generation of ROS and RNS, leading to increased oxidative stress. In the absence of an appropriate compensatory response from endogenous antioxidant defences, the system becomes overwhelmed (redox imbalance), leading to the activation of stress-sensitive signalling pathways, such as NF-kB, p38 mitogen-activated protein kinase (MAPK), Jun

kinases (JNK), stress-activated protein kinases (SAPK), PKC, AGE, receptor for advanced glycosylation end-products (RAGE) and sorbitol. Hence, oxidative stress, induced by high glucose load followed by activation of other pathways cause cellular damage, and are ultimately responsible for the long-term complications of diabetes [54].

TREATMENT OF INSULIN RESISTANCE

The current non-pharmacological avenues of increasing insulin sensitivity in T2DM are exercise and diet, with biguanides, sulphonylureas and thiazolidinediones as the mainstays of pharmacological treatment. Insulin is usually reserved for use in secondary failure of oral agents and in diabetic hyperglycaemic emergencies.

ALA has shown therapeutic potential in the area of insulin resistance [55]. A four-week placebo-controlled euglycaemic glucose-clamp study using oral ALA at 600 mg daily in patients with T2DM showed an improvement of insulin-stimulated glucose disposal of 27% compared to placebo [56]. This improvement in insulin sensitivity was also replicated in a more recent study using oral ALA at 600 mg twice a day [57]. Being a powerful antioxidant, ALA has shown some promise as a safe alternative in reducing insulin resistance through augmenting insulin stimulated glucose disposal, inhibiting inflammatory cytokines and decreasing ROS production [58-60].

Another important action of ALA is on the expression of AMP-activated protein kinase (AMPK) in the hypothalamus and peripheral tissues. AMPK functions as a fuel sensor in the cell and is activated when cellular energy is depleted reflected by an increase in AMP:ADP ratio [61]. AMPK activation in skeletal muscle improves insulin sensitivity by increasing glucose uptake and fatty acid oxidation. It does this by promoting glucose transport through the translocation of GLUT-4 to the plasma membrane in an insulin-independent manner [22]. Insulin sensitivity is also improved through the reduction in triglyceride accumulation in skeletal muscle [54]. This occurs as a result of AMPK phosphorylating, and thus inactivating acetyl-CoA carboxylase 2 (ACC-2 or β) leading to a decrease in malonyl-coenzyme A (malonyl-CoA). Decreased malonyl-CoA would thus cause an increase in long fatty chain acid oxidation [62, 63]. Interestingly, AMPK is also responsible for mediating the effects of the oral hypoglycaemic agents such as metformin and peroxisome proliferator-activated receptors (PPAR) gamma agonists [64]. Finally, ALA exerts potent anti-obesity effects by decreasing hypothalamic AMPK activity and reducing food intake thereby causing profound weight loss *in vivo* [65].

TREATMENT OF DIABETIC NEUROPATHY

One of the commonest complications developing in at least one-third of all patients is diabetic neuropathy (DN). Diabetic neuropathy is a dynamic and chronic degenerative disorder which commonly manifests as a distal symmetrical polyneuropathy. It has multiple interactive pathogenetic mechanisms including perfusion abnormalities and oxidative stresses [66], which explains the difficulty in designing efficacious and biologically-meaningful therapies [67]. The first step in the long-term treatment of DN is to aim for a stable and optimal glycaemic control. Common pharmacological

treatment of DN includes the use of amitriptyline, anticonvulsants and opioids, which have been shown to alleviate the varied spectrum of symptoms like burning, paraesthesias and pain, but at the significant expense of adverse effects, and effective pain relief is reported to be achieved in less than half of patients with chronic neuropathic pain [68].

Several newer agents have been shown to be efficacious in randomized controlled trials, but with the exception of duloxetine and pregabalin, none has been specifically licensed for the management of painful DN [69, 70]. Even so, there was a higher drop rate in patients on duloxetine due to adverse events (13.9% vs 7.2%), suggesting that it should be used with similar caution like the other anti-depressants [71].

From the understanding so far of the conceptual framework of the multi-factorial pathogenesis and treatment of DN from animal studies and cell cultures, there is still no long term effective pathogenetic treatment for DN to date, although a few drugs are currently being studied in clinical trials. ALA, however, has shown much promise [72], and has already been used as treatment for DN in some European countries like Germany.

Given the lack of consensus concerning the appropriate first line treatment of neuropathic pain, due to a lack of head to head comparisons which limits any definitive conclusions in this area, clinicians must therefore make decisions regarding the care of individual patients using randomized trials with positive outcome for each agent [69, 72, 73].

One of the possible beneficial effects of ALA is on the improvement of microcirculation as it has been shown to increase perfusion reserve on demand as demonstrated by a decrease in the time to peak capillary blood cell velocity during post-occlusive hyperemia both orally (12.6 ± 3.1 s vs 35.4 ± 10.9 s, $p < 0.05$) and intravenously (11.74 ± 4.4 s vs 21.9 ± 5.0 s, $p < 0.05$) [74]. The authors suggested that this likely reflected an improvement of the pathophysiology of the nerves rather than an actual regeneration of nerve endings. Moreover, oral ALA 800mg/day for 4 months was also shown to have favourable effects on cardiac autoneuropathy in T2DM patients [75].

Several European studies have demonstrated the benefits of ALA in symptomatic polyneuropathy. In the (Alpha Lipoic Acid in Diabetic Neuropathy I) ALADIN-I study, intravenous ALA was shown to improve symptom scores [76] while the ALADIN-II study demonstrated meaningful dose-dependent improvements in neuropathic functions with oral ALA for a two-year duration, reflected as an improvement in sural sensory nerve conduction velocity ($+3.8 \pm 4.2$ m/s vs -0.1 ± 4.8 m/s, $p < 0.05$), sural sensory nerve action potential ($+0.3 \pm 1.4$ V vs -0.7 ± 1.5 V, $p < 0.05$) and in tibial motor nerve conduction velocity ($+1.2 \pm 3.8$ m/s vs -1.5 ± 2.9 m/s, $p < 0.05$) [77].

In the SYDNEY Trial, oral treatment with ALA 600mg thrice daily for three weeks or intravenous ALA 600mg infused five times per week were shown to improve symptoms on the Total Symptom Score (TSS), Hamburg Pain Adjective List (HPAL), Neuropathy Disability Score (NDS) and Neuropathic Impairment Score (NIS) as compared to placebo [78, 79].

Two multi-centre studies are now underway to determine the role of ALA in the prevention and treatment of diabetic neuropathy. The Neurological Assessment of Thioctic Acid in Neuropathy I (NATHAN I) Study, is a pivotal long-term trial using oral ALA conducted in North America and Europe spanning four years and is aimed at slowing the progression of diabetic polyneuropathy, while NATHAN II addresses the use of an intravenous agent for symptomatic relief of painful neuropathy [80].

ADVERSE EFFECTS

In terms of side effect profile, the SYDNEY-2 trial demonstrated a dose-dependent increase in nausea, vomiting, and vertigo with increasing doses of ALA from 600mg up to a maximum of 1800mg. Hence, an oral dose of 600mg once daily was recommended by the investigators in order to provide the optimal risk-benefit ratio [81].

The risk of hypoglycaemia with the use of ALA has been attributed to an autoimmune process called Insulin Autoimmune Syndrome (IAS), in addition to potentiating the effects of other hypoglycaemic agents like insulin therapy [82, 83]. IAS (also known as Hirata's disease) was first described in 1970 and is a relatively rare cause of hypoglycaemia [84]. It is characterized by the production of auto-antibodies to insulin in individuals who have not previously been exposed to insulin. Another risk factor for the development of this complication is the use of sulfhydryl drugs like methimazole, mercaptopropionyl glycine and glutathione. It is therefore believed that IAS is a form of drug-induced autoimmunity as a result of drug-drug interaction because ALA, being a disulphide derivative, has the ability to also produce a sulfhydryl group when reduced *in vivo* [85]. Finally, the risk is also higher in East Asians and certain North American natives, among whom the prevalence of HLA alleles that confer predisposition to this syndrome is relatively high [86].

CONCLUSION

New headways are currently being forged in the understanding of the pathogenesis of T2DM and its complications. Specific targeted therapy by ALA has been shown to be both an attractive and relevant adjuvant pharmacotherapy for DN [87]. Given the growing evidence of increased oxidative and inflammatory load implicated in T2DM, treatment paradigms may have to shift to target these stressors with the use of compounds like ALA. Adequately powered randomized placebo controlled trials are urgently needed to establish its role in the treatment of hyperglycemia and DN in T2DM. For example, given that ALA is rapidly metabolized and excreted with peak plasma concentrations lasting less than an hour, the clinical and long term value of ALA in improving glycaemic control as reflected by the glycated haemoglobin (HbA1c) levels need to be established [27].

Although a change in lifestyle is the best preventive and therapeutic approach, ALA may prove to be an important part of the therapeutic armamentarium in combating the worldwide epidemic of diabetes, along with other established arms of treatment. In conclusion, the use of ALA represents a potential and attractive pharmacological target in the treatment of T2DM by modulating the signal transduction pathways in insulin resistance and antagonizing the oxidative

and inflammatory stresses, which are major players in the pathogenesis of this disorder.

ABBREVIATIONS

ACC-2 or β	=	Acetyl-CoA Carboxylase 2
AGEs	=	Advanced Glycated End products
ALADIN-I	=	Alpha Lipoic Acid in Diabetic Neuropathy I
ALADIN-II	=	Alpha Lipoic Acid in Diabetic Neuropathy II
ALA	=	Alpha Lipoic Acid
AMPK	=	AMP-activated Protein Kinase
CCl ₄	=	Carbon Tetrachloride
DCCT	=	Diabetes Control and Complications Trial
DHLA	=	Dihydrolipoic Acid
DN	=	Diabetic Neuropathy
ESRF	=	End-Stage Renal Failure
FFA	=	Free Fatty Acid
GLUT	=	Glucose Transporter
HPAL	=	Hamburg Pain Adjective List
IAS	=	Insulin Autoimmune Syndrome
IKK β	=	Inhibitor of nuclear factor Kappa b Kinase
IR	=	Insulin Receptors
JNK	=	Jun Kinases
MAPK	=	Mitogen-Activated Protein Kinase
Malonyl-CoA	=	Malonyl-Coenzyme A
NAD ⁺	=	Nicotinamide Adenine Dinucleotide
NADPH	=	Nicotinamide Adenine Dinucleotide Phosphate
NATHAN I	=	Neurological Assessment of Thioctic Acid in Neuropathy I
NATHAN II	=	Neurological Assessment of Thioctic Acid in Neuropathy II
NDS	=	Neuropathy Disability Score
NF- κ B	=	Nuclear Factor-Kappa B
NIS	=	Neuropathic Impairment Score
PKC	=	Protein Kinase C
PKC ζ	=	Protein Kinase C ζ
PPAR	=	Peroxisome Proliferator-Activated Receptor
R-LA	=	R-enantiomer of Lipoic Acid
RAGE	=	Receptor for Advanced Glycosylation End-product
ROS	=	Reactive Oxygen Species

RNS	=	Reactive Nitrogen Species
S-LA	=	S-enantiomer of Lipoic Acid
SAPK	=	Stress-Activated Protein Kinase
TNF- α	=	Tumour Necrosis Factor-Alpha
TSS	=	Total Symptom Score
T2DM	=	Type 2 Diabetes Mellitus
UKPDS	=	United Kingdom Prospective Diabetes Study

REFERENCES

- Reed, L.J. (2001) A trail of research from lipoic acid to alpha-keto acid dehydrogenase complexes. *J. Biol. Chem.*, **276**, 38329-38336.
- Smith, A.R.; Shenvi, S.V.; Widlansky, M.; Suh, J.H. and Hagen, T.M. (2004) Lipoic acid as a potential therapy for chronic diseases associated with oxidative stress. *Curr. Med. Chem.*, **11**, 1135-1146.
- Lester, P.; Peter, R.; Tritschler, H.J.; King, G.L.; Azzi, A. (2000) *Antioxidants in Diabetes Management*. New York NY: Marcel Dekker, Inc.
- Bustamante, J.; Lodge, J.K.; Marcocci, L.; Tritschler, H.J.; Packer, L. and Rihl, B.H. (1998) Alpha-lipoic acid in liver metabolism and disease. *Free Radic. Biol. Med.*, **24**, 1023-1039.
- Jakuss, V. (2000) The role of free radicals, oxidative stress and antioxidant systems in diabetic vascular disease. *Bratisl. Lek. Listy.*, **101**, 541-551.
- Halliwell, B. and Gutteridge, J.M.C.; Eds. (1999) *Free Radicals in Biology and Medicine*, Oxford University Press: New York.
- Sies, H. Ed. (1991) *Oxidative Stress II: Oxidants and Antioxidants*, Academic Press: London.
- Harman, D. (1981) The aging process. *Proc. Natl. Acad. Sci. USA*, **78**, 7124-7128.
- Harman, D. (1956) Aging: a theory based on free radical and radiation chemistry. *Gerontology*, **11**, 298-300.
- Ozawa T. (1997) Genetic and functional changes in mitochondria associated with aging. *Physiol. Rev.*, **77**, 425-464.
- Lenaz, G. (1998) Role of mitochondria in oxidative stress and aging. *Biochim. Biophys. Acta*, **1366**, 53-67.
- Chance, B.; Sies, H. and Boveris, A. (1979) Hydroperoxide metabolism in mammalian organs. *Physiol. Rev.*, **59**, 527-605.
- Suh, J.H.; Shenvi, S.V.; Dixon, B.M.; Liu, H.; Jaiswal, A.K.; Liu, R.M. and Hagen, T.M. (2004) Decline in transcriptional activity of Nrf2 causes age-related loss of glutathione synthesis, which is reversible with lipoic acid. *Proc. Natl. Acad. Sci. USA*, **101**, 3381-3386.
- Suh, J.H.; Wang, H.; Liu, R.M.; Liu, J. and Hagen, T.M. (2004) (R)-alpha-lipoic acid reverses the age-related loss in GSH redox status in post-mitotic tissues: evidence for increased cysteine requirement for GSH synthesis. *Arch. Biochem. Biophys.*, **423**, 126-135.
- Lester, P. and Eric, H. (1995) Witt and Hans Jürgen Tritschler alpha lipoic acid as a biological antioxidant. *Free Radic. Biol. Med.*, **19**, 227-250.
- Kozlov, A.V.; Gille, L.; Staniek, K. and Nohl, H. (1999) Dihyrolipoic acid maintains ubiquinone in the antioxidant active form by two-electron reduction of ubiquinone and one-electron reduction of ubisemiquinone. *Arch. Biochem. Biophys.*, **363**, 148-154.
- Jones, W.; Li, X.; Qu, Z.C.; Perriott, L.; Whitesell, R.R. and May, J.M. (2002) Uptake, recycling, and antioxidant actions of alpha-lipoic acid in endothelial cells. *Free Radic. Biol. Med.*, **33**, 83-93.
- Ou, P.; Tritschler, H.J. and Wolff, S.P. (1995) Thioctic (lipoic) acid: a therapeutic metal-chelating antioxidant? *Biochem. Pharmacol.*, **50**, 123-126.
- Suh, J.H.; Zhu, B.Z.; deSzoeko, E.; Frei, B. and Hagen, T.M. (2004) Dihyrolipoic acid lowers the redox activity of transition metal ions but does not remove them from the active site of enzymes. *Redox Rep.*, **9**, 57-61.
- Suh, J.H.; Moreau, R.; Heath, S.H. and Hagen, T.M. (2005) Dietary supplementation with (R)-alpha-lipoic acid reverses the age-related accumulation of iron and depletion of antioxidants in the rat cerebral cortex. *Redox Rep.*, **10**, 52-60.
- Biewenga, G.P.; Haenen, G.R. and Bast, A. (1997) The pharmacology of the antioxidant lipoic acid. *Gen. Pharmacol.*, **29**, 315-331.
- Konrad, D.; Somwar, R.; Sweeney, G.; Yaworsky, K.; Hayashi, M.; Ramlal, T. and Klip, A. (2001) The antihyperglycemic drug alpha-lipoic acid stimulates glucose uptake via both GLUT4 Translocation and GLUT4 Activation. *Diabetes*, **50**, 1464-1471.
- Konrad, D. (2005) Utilization of the insulin-signaling network in the metabolic actions of alpha-lipoic acid-reduction or oxidation? *Antioxid. Redox Signal*, **7**, 1032-1039.
- Packer, L. (1998) Alpha-Lipoic acid: a metabolic antioxidant which regulates NF-kappa B signal transduction and protects against oxidative injury. *Drug Metab. Rev.*, **30**, 245-275.
- Mark, F.; McCarty, B.A. and Keith, I. (2006) Block, preadministration of high-dose salicylates, suppressors of NF- κ B activation, may increase the chemosensitivity of many cancers: an example of proapoptotic signal. *Modulation Ther. Integr. Cancer Ther.*, **5**, 252-268.
- Zhang, W.J. and Frei B. (2001) Alpha-lipoic acid inhibits TNF-alpha-induced NF-kappaB activation and adhesion molecule expression in human aortic endothelial cells. *FASEB J.*, **15**, 2423-2432.
- Hermann, R.; Niebch, G.; Borbe, H.O.; Fieger-Büschges H.; Ruus, P.; Nowak, H.; Riethmüller-Winzen, H.; Peukert, M. and Blume H. (1996) Enantioselective pharmacokinetics and bioavailability of different racemic alpha-lipoic acid formulations in healthy volunteers. *Eur. J. Pharm. Sci.*, **4**, 167-174.
- Gleiter, C.H.; Schug, B.S.; Hermann, R.; Elze, M.; Blume, H.H. and Gundert-Remy, U. (1996) Influence of food intake on the bioavailability of thioctic acid enantiomers. *Eur. J. Clin. Pharmacol.*, **50**, 513-514.
- Uma, S. and Ishwarlal, J. (2008) Alpha-lipoic acid supplementation and diabetes. *Nutr. Rev.*, **66**, 646-657.
- Borcea, V.; Nourooz-Zadeh, J.; Wolff, S.P.; Klevesath, M.; Hofmann, M.; Ulrich, H.; Wahl, P.; Ziegler, R.; Tritschler, H.; Halliwell, B. and Nawroth, P.P. (1999) Alpha Lipoic acid decreases oxidative stress even in diabetic patients with poor glycemic control and albuminuria. *Free Radic. Biol. Med.*, **26**, 1495-1500.
- Teichert, J.; Hermann, R.; Ruus, P. and Preiss, R. (2003) Plasma kinetics, metabolism, and urinary excretion of alpha-lipoic acid following oral administration in healthy volunteers. *J. Clin. Pharmacol.*, **43**, 1257-1267.
- Teichert, J.; Tuemmers, T.; Achenbach, H.; Preiss, C.; Hermann, R.; Ruus, P. and Preiss, R. (2005) Pharmacokinetics of alpha-lipoic acid in subjects with severe kidney damage and end-stage renal disease. *J. Clin. Pharmacol.*, **45**, 313-328.
- Juanita, B.A.; John, K.; Lodge, A.; Lucia, M.A.; Hans, J.; Tritschler, B.; Lester, P.A.; Bertrand, H. and Rihl, A. (1998) α -Lipoic acid in liver metabolism and disease. *Free Radic. Biol. Med.*, **24**, 1023-1039.
- Bartter, F.C.; Berkson, B.M.; Galleli, J. and Hiranaka, P. (1980) Thioctic Acid in the Treatment of Poisoning with Alpha-Amanitin. In: Faulstich, H.; Kommerell, B.; Wieland, T. Eds. *Amanita Toxins and Poisoning*. Baden-Baden Germany: Verlag, Gerhard Witzstroch; pp. 197-202.
- Berkson, B.M. (1999) A conservative triple antioxidant approach to the treatment of hepatitis C. combination of alpha lipoic acid (thioctic acid), silymarin, and selenium: three case histories. *Med. Klin (Munich)*, **94**, 84-89.
- Evans, J.L.; Youngren, J.F. and Goldfine, I.D. (2004) Effective treatments for insulin resistance: Trim the fat and douse the fire. *Trends Endocrinol. Metab.*, **15** (9), 425-431.
- Hotamisligil, G.S. and Spiegelman, B.M. (1994) Tumor necrosis factor alpha: a key component of the obesity-diabetes link. *Diabetes*, **43**, 1271-1278.
- Ceriello, A. and Motz, E. (2004) Is oxidative stress the pathogenic mechanism underlying insulin resistance, diabetes, and cardiovascular disease? The common soil hypothesis revisited. *Arterioscler. Thromb. Vasc. Biol.*, **24**, 816-823.
- Diabetes Study (UKPDS) Group. Intensive blood-glucose control with sulphonylureas or insulin compared with conventional treatment and risk of complications in patients with type 2 diabetes (UKPDS 33). (1998) UK Prospective. *Lancet*, **352**, 837-853.

- [40] Douglas, A.; Greene, S.A. and Littmer, A.A.F. (1988) Are disturbances of sorbitol, phosphoinositide, and Na⁺-K⁺-ATPase regulation involved in pathogenesis of diabetic neuropathy? *Perspect. Diabetes*, **37**, 688-693.
- [41] Greene, D.A.; DeJesus, P.V. and Winegrad, A.I. (1975) Effects of insulin and dietary myo-inositol on impaired peripheral motor nerve conduction velocity in acute streptozotocin diabetes. *J. Clin. Invest.*, **55**, 1326-1336.
- [42] Mayer, J.H. and Tomlinson, D.R. (1983) Prevention of defects of axonal transport and nerve conduction velocity by oral administration of myo-inositol or an aldose reductase inhibitor 11- streptozotocin-dlabetc rats. *Diabetologia*, **25**, 433-438.
- [43] Stevens, M.J.; Lattimer, S.A.; Kamijo, M.; Van Huysen, C.; Sima, A.A. and Greene, D.A. (1993) Osmotically-induced nerve taurine depletion and the compatible osmolyte hypothesis in experimental diabetic neuropathy in the rat. *Diabetologia*, **36**, 608-614.
- [44] Cameron, N.E.; Eaton, S.E.; Cotter, M.A. and Tesfaye, S. (2001) Vascular transport and metabolic interactions in the pathogenesis of diabetic neuropathy. *Diabetologia*, **44**, 1973-1988.
- [45] Diñçer, Y.; Akçay, T.; İlkova, H.; Alademir, Z. and Özbay, G. (2003) DNA damage and antioxidant defense in peripheral leukocytes of patients with Type I diabetes mellitus. *Mutat. Res.*, **527**, 49-55.
- [46] Erika, S.; József, P.; János, F. and Anikó, S. (1999) Alterations in enzymatic antioxidant defence in diabetes mellitus a rational approach. *Postgrad. Med. J.*, **75**, 13-17.
- [47] Andreeva-Gateva, P.; Popova, D. and Orbetsova, V. (2001) Antioxidant parameters in metabolic syndrome -- a dynamic evaluation during oral glucose tolerance test. *Vutr. Boles.*, **33**, 48-53.
- [48] Oberley, L.W. (1988) Free radicals and diabetes. *Free Radic. Biol. Med.*, **5**, 113-124.
- [49] Rösen, P.; Nawroth, P.P.; King, G.; Möller, W.; Tritschler, H.J. and Packer, L. (2001) The role of oxidative stress in the onset and progression of diabetes and its complications: a summary of a Congress Series sponsored by UNESCO-MCBN, the American Diabetes Association and the German Diabetes Society. *Diabetes Metab. Res. Rev.*, **17**, 189-212.
- [50] Bonnefont-Rousselot, D. (2002) Glucose and reactive oxygen species. *Curr. Opin. Clin. Nutr. Metab. Care*, **5**, 561-568.
- [51] Brownlee, M. (2001) Biochemistry and molecular cell biology of diabetic complications. *Nature*, **414**, 813-820.
- [52] Varma, S.D.; Devamanoharan, P.S. and Ali, A.H. (1997) Formation of advanced glycation end (AGE) products in diabetes: prevention by pyruvate and α -ketoglutarate. *Mol. Cell. Biochem.*, **171**, 23-28.
- [53] Dorota, Z. and Bogna, W.-W. (2005) Hyperglycaemia and Inflammation are culprits of late diabetic complications. *Arch. Med. Sci.*, **1**, 115-118.
- [54] Evans, J.L.; Goldfine, I.D.; Maddux, B.A. and Grodsky, G.M. (2002) Oxidative stress and stress-activated signaling pathways: a unifying hypothesis of type 2 diabetes. *Endocr. Rev.*, **23**, 599-622.
- [55] Jacob, S.P.; Ruus, R.; Hermann, H.J.; Tritschler, E.; Maerker, W.; Renn, H.J.; Augustin, G.J. and Rett, K. (1999) Oral administration of rac-alpha-lipoic acid modulates insulin sensitivity in patients with type-2 diabetes mellitus: a placebo controlled pilot trial. *Free Radic. Biol. Med.*, **27**(3-4), 309-314.
- [56] Jacob, S.; Ruus, P.; Hermann, R.; Tritschler, H.J.; Maerker, E.; Renn, W.; Augustin, H.J.; Dietze, G.J. and Rett, K. (1999) Oral administration of RAC-alpha-lipoic acid modulates insulin sensitivity in patients with type-2 diabetes mellitus: a placebo-controlled pilot trial. *Free Radic. Biol. Med.*, **27**(3-4), 309-314.
- [57] Kamenova, P. (2006) Improvement of insulin sensitivity in patients with type 2 diabetes mellitus after oral administration of alpha-lipoic acid. *Hormones (Athens)*, **5**, 251-258.
- [58] Midaoui, A.E.L. and Jacques de Champlain. (2002) Prevention of hypertension, insulin resistance and oxidative stress by alpha-lipoic acid. *Hypertension*, **39**, 303-307.
- [59] Jacob, S.; Henriksen, E.J.; Schiemann, A.L.; Simon, I.; Clancy, D.E.; Tritschler, H.J.; Jung, W.I.; Augustin, H.J. and Dietze, G.J. (1995) Enhancement of glucose disposal in patients with type 2 diabetes by alpha-lipoic acid. *Arzneimittelforschung*, **45**, 872-874.
- [60] Karen, S. and William, C. (2002) Gong. Natural products used for Diabetes. *J. Am. Pharm. Assoc.*, **42**, 217-226.
- [61] Blerina, K.; Marco, B.; Rutter, G.A.; Grossman, A.B. and Korbonits, M. (2004) Expanding role of AMPK in endocrinology. *Trends Endocrinol. Metab.*, **17**, 58.
- [62] Kahn, B.B.; Alquier, T.; Carling, D. and Hardie, D.G. (2005) AMP-activated protein kinase: ancient energy gauge provides clues to modern understanding of metabolism. *Cell. Metab.*, **1**, 15-25.
- [63] Richter, E.A.; Derave, W. and Wojtaszewski, J.F. (2001) Glucose, exercise and insulin: emerging concepts. *J. Physiol.*, **535**, 313-322.
- [64] Hardie, D.G.; Scott, J.W.; Pan, D.A. and Hudson, E.R. (2003) Management of cellular energy by the AMP-activated protein kinase system. *FEBS Lett.*, **546**, 113-120.
- [65] Kola, B.; Boscaro, M.; Rutter, G.A.; Grossman, A.B. and Korbonits, M. (2006) Expanding role of AMPK in endocrinology. *Trends Endocrinol. Metab.*, **17**, 205-215.
- [66] Cameron, N.E. and Cotter, M.A. (1997) Metabolic and vascular factors in the pathogenesis of diabetic neuropathy. *Diabetes*, **46**, S31-S37.
- [67] Sima, A.A.F. (2001). Diabetic neuropathy: pathogenetic background, current and future therapies. *Expert Rev. Neurother.*, **1**, 225-238.
- [68] Kenneth, C. Jackson II. (2006) Pharmacotherapy for Neuropathic Pain. *Pain Practice*. Blackwell Publishing. Volume 6 (Issue 1) pp. 27-33.
- [69] Bouton, A.J.; Viniki, A.I.; Arezzo, J.C.; Bril, V.; Feldman, E.L.; Freeman, R.; Malik, R.A.; Maser, R.E.; Sosenko, J.M. and Ziegler, D. Diabetic Neuropathies A statement by the American Diabetes Association. *Diabetes Care*, **28**, 956-962. 2005.
- [70] Rosenstock, J.; Tuchman, M.; LaMoreaux, L. and Sharma, U. (2004) Pregabalin for the treatment of painful diabetic neuropathy: a double-blind, placebo-controlled trial. *Pain*, **110**, 628-638.
- [71] Goldstein, D.J. (2006) Duloxetine in the treatment of diabetic peripheral neuropathic pain. *Future Neurol.*, **1**, 701-711.
- [72] Vinik, A. and Mehrabyan, A. (2004) Diabetic neuropathies. *Med. Clin. N. Am.*, **88**, 947-999.
- [73] Misha-Miroslav, B. and Serra, J. (2004) Pharmacologic management part I: better-studied neuropathic pain diseases. *Pain Med.*, **5**, S28-S47.
- [74] Haak, E.; Usadel, K.H.; Kusterer, K.; Amini, P.; Frommeyer, R.; Tritschler, H.J. and Haak, T. (2000) Effects of alpha-lipoic acid on microcirculation in patients with peripheral diabetic neuropathy. *Exp. Clin. Endocrinol. Diabetes*, **108**, 168-174.
- [75] Ziegler, D.; Schatz, H.; Conrad, F.; Gries, F.A.; Ulrich, H. and Reichel, G. (1997) Effects of treatment with the antioxidant alpha-lipoic acid on cardiac autonomic neuropathy in NIDDM patients. A 4-month randomized controlled multi-centre trial (DEKAN Study). *Deutsche Kardiale Autonome Neuropathie. Diabetes Care*, **20**, 1918-1920.
- [76] Ziegler, D.; Hanefeld, M.; Ruhnau, K.J.; Meissner, H.P.; Lobisch, M.; Schütte, K. and Gries, F.A. (1995) Treatment of symptomatic diabetic peripheral neuropathy with the anti-oxidant alpha-lipoic acid. A 3-week multicentre randomized controlled trial (ALADIN Study). *Diabetologia*, **38**, 1425-1433.
- [77] Reljanovic, M.; Reichel, G.; Rett, K.; Lobisch, M.; Schuette, K.; Möller, W.; Tritschler, H.J. and Mehnert, H. (1999) Treatment of diabetic polyneuropathy with the antioxidant thioctic acid (alpha-lipoic acid): a two year multicenter randomized double-blind placebo-controlled trial (ALADIN II): alpha lipoic acid in diabetic neuropathy. *Free Radic. Res.*, **31**, 171-179.
- [78] Ametov, A.S.; Barinov, A.; Dyck, P.J.; Hermann, R.; Kozlova, N.; Litchy, W.J.; Low, P.A.; Nehrdich, D.; Novosadova, M.; O'Brien, P.C.; Reljanovic, M.; Samigullin, R.; Schuette, K.; Stokov, I.; Tritschler, H.J.; Wessel, K.; Yakhno, N.; Ziegler, D. and SYDNEY Trial Study Group. (2003) The sensory symptoms of diabetic polyneuropathy are improved with alpha-lipoic acid: the SYDNEY trial. *Diabetes Care*, **26**, 770-776.
- [79] Ruhnau, K.J.; Meissner, H.P.; Finn, J.R.; Reljanovic, M.; Lobisch, M.; Schütte, K.; Nehrdich, D.; Tritschler, H.J.; Mehnert, H. and Ziegler D. (1999) Effects of 3-week oral treatment with the antioxidant thioctic acid (α -lipoic acid) in symptomatic diabetic polyneuropathy. *Diabet. Med.*, **6**, 1040-1043.
- [80] Ziegler, D.; Reljanovic, M.; Mehnert, H. and Gries, F.A. (1999) Alpha-lipoic acid in the treatment of diabetic polyneuropathy in Germany: current evidence from clinical trials. *Exp. Clin. Endocrinol. Diabetes*, **107**, 421-430.

- [81] Ziegler, D.; Ametov, A.; Barinov, A.; Dyck, P.J.; Gurieva, I.; Low, P.A.; Munzel, U.; Yakhno, N.; Raz, I.; Novosadova, M.; Maus, J. and Samigullin, R. (2006) Oral treatment with alpha-lipoic acid improves symptomatic diabetic polyneuropathy: the SYDNEY 2 trial. *Diabetes Care*, **29**, 2365-2370.
- [82] Takeuchi, Y.; Miyamoto, T.; Kakizawa, T.; Shigematsu, S. and Hashizume, K. (2007) Insulin autoimmune syndrome possibly caused by alpha lipoic acid. *Intern. Med.*, **46**(5), 237-239.
- [83] Furukawa, N.; Miyamura, N.; Nishida, K.; Motoshima, H.; Taketa, K. and Araki, E. (2007) Possible relevance of alpha lipoic acid contained in a health supplement in a case of insulin autoimmune syndrome. *Diabetes Res. Clin. Pract.*, **75**(3), 366-367.
- [84] Yasuko, I. and Yukimasa, U. (1999) HIRATA. Insulin autoimmune syndrome (IAS, Hirata disease)1. *Ann. Med. Interne (Paris)*., **150**, 245-253.
- [85] Yoshihiko, I.; Takeshi, O.; Yoko, O.; Tatsuo, I.; Yushi, H.; Kensuke, F.; Kazuhiko, S.; Wataru, O. and Masato, K. (2007) Alpha-lipoic and insulin autoimmune syndrome. *Diabetes Care*, **30**, 2240-2241.
- [86] Uchigata, Y.; Hirata, Y.; Omori, Y.; Iwamoto, Y. and Tokunaga, K. (2006) Worldwide differences in the incidence of insulin autoimmune syndrome (Hirata disease) with respect to the evolution of HLA-DR4 alleles. *Hum. Immunol.*, **61**, 154-157.
- [87] Ziegler, D. (2004) Thioctic acid for patients with symptomatic diabetic polyneuropathy: a critical review. *Treat. Endocrinol.*, **3**, 173-189.

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